
Review Article


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CHEMICAL CONSTITUENTS AND PHARMACOLOGICAL ACTIVITIES OF *AMMI MAJUS* AND *AMMI VISNAGA*. A REVIEW

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Abstract

Ammi species belong to the family Umbellifereae, contained bioactive compounds (mainly coumarins and flavonoids) of important biological activities. *Ammi majus* fruit contained amorphous glucoside 1%, tannin 0.45%, oleoresin 4.76%, acrid oil 3.2%, fixed oil 12.92%, proteins 13.83% and cellulose 22.4%. However, the major constituents of *Ammi majus* are the furanocoumarins, which included xanthotoxin (methoxsalen, 8-methoxypsoralen, ammoidin, up to 1.15%), imperatorin (ammidin, up to 0.75%) and bergapten (heraclin, majudin, 5-methoxypsoralen, up to 1.88%), marmesin 0.25%, isoimperatorin 0.01%, heraclenin 0.07% and isopimpinellin 0.01%. *Ammi visnaga* contained -pyrones (furanochromone up to 4%), the principal compounds being khellin (0.3–1.2%), visnagin (0.05–0.30%), khellinol, ammiol, khellol and khellinin. *Ammi visnaga* also contained fixed oils (up to 18%) and coumarins (0.2–0.5%), the main one being the pyranocoumarin visnadin (0.3%). The previous pharmacological studies showed that *Ammi majus* was used effectively in the treatment of psoriasis, vitiligo and tinea versicolor. Its furocoumarins have bactericidal, fungicidal, insecticidal, larvicidal, moluscicidal, nematocidal, ovicidal, viricidal and herbicidal activities. *Ammi visnaga* was also used effectively for the treatment of vitiligo. It exerted a wide range of antibacterial activity and induced smooth muscle relaxant effects especially vascular smooth muscle. The present review will highlight the chemical constituents and the pharmacological and therapeutic effects of *Ammi majus* and *Ammi visnaga*.

Keywords: *Ammi majus*, *Ammi visnaga*, Furanocoumarins, Flavonoids, Vitiligo, Psoriasis.

Introduction

Plants are a valuable source of a wide range of secondary metabolites, which are used as pharmaceuticals, agrochemicals, flavors, fragrances, colors, biopesticides and food additives. Medicinal plants are the Nature's gift to human beings to help them pursue a disease-free healthy life. Plants have been used as drugs by humans since thousands of years ago. As a result of accumulated experience from the past generations, today, all the world's cultures have an extensive knowledge of herbal

medicine. *Ammi* species, belong to the family Umbellifereae, contained bioactive compounds (mainly coumarins and flavonoids) of important biological activities. *Ammi majus* is indigenous to Egypt and it grows in the Nile Valley, especially in Behira and Fayoom. It is also found in the basin of the Mediterranean Sea, West Africa, in some regions of Iran and the mountains of Kohaz. *Ammi visnaga* is distributed in North Africa, Europe, Eastern Mediterranean region, South western Asia,

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North America, Argentina, Chile, Mexico, and Atlantic Islands. In Iraq, *Ammi majus* usually found in fields and gardens and by the side of channels, often as weed of cultivation. It is collected from Kut, Baghdad, Hawija and many other areas, while *Ammi visnaga* is distributed in Erbil, Mousl, Baghdad, Sulaimania and Kirkuk in north of Iraq¹⁻³.

The dried ripe fruits of *Ammi majus* were used traditionally for the treatment of skin disorders, psoriasis and vitiligo. It was used as an emmenagogue to regulate menstruation, as a diuretic, and for treatment of leprosy, kidney stones, and urinary tract infections⁴⁻⁹. While, *Ammi visnaga* was used traditionally in the treatment of mild anginal symptoms, as supportive treatment for mild obstruction of the respiratory tract in asthma, bronchial asthma or spastic bronchitis, and postoperative treatment of conditions associated with the presence of urinary calculi. It was also used for the treatment of gastrointestinal cramps, as diuretic, for painful menstruation and as an emmenagogue to regulate menstruation¹⁰. The aim of this study is to highlight the chemical constituents and the pharmacological and therapeutic effects of *Ammi majus* and *Ammi visnaga*

I-Ammi majus

Synonym: *Apium ammi*

Common names : English : Bishop's weed, Greater Ammi; Arabic: Khella shaitani , Khella bariah

Traditional use: The fruits were used for the treatment of skin disorders, psoriasis and vitiligo. It was also used as an emmenagogue to regulate menstruation, as a diuretic, and for treatment of leprosy, kidney stones, and urinary tract infections⁴⁻⁹.

Physicochemical properties¹¹: moisture content : (loss on drying at 105°C) - 5.25% w/w , total ash content of powdered drug - 7.00% w/w , water soluble ash - 5.35% w/w , and acid insoluble ash - 0.86 % w/w . Extractive value in different solvents %: Acetone - 6.00 w/w, absolute alcohol - 3.50 w/w, chloroform - 1.75 w/w, methanol - 7.85 w/w, petroleum ether (60-80) - 1.20 w/w, and water - 17.35 w/w.

Chemical constituents

Ammi majus fruits contained amorphous glucoside 1%, tannin 0.45%, oleoresin 4.76%, acrid oil 3.2%, fixed oil 12.92%, proteins 13.83% and cellulose

22.4%³. The major constituents of *Ammi majus* are the furanocoumarins, which included xanthotoxins (methoxsalen, 8-methoxypsoralen , ammoidin , up to 1.15%), imperatorin (ammidin, up to 0.75%) and bergaptens (heraclin, majudin, 5-methoxypsoralen , up to 1.88%) , marmesin 0.25% , isoimperatorin ,0.01%, heraclenin 0.07% and isopimpinellin 0.01%¹²⁻²¹. Selim and Ouf isolated two coumarins from the aerial parts of the Egyptian *Ammi majus* L. , 6- hydroxy-7-methoxy-4 methyl coumarin and 6-hydroxy-7-methoxy coumarin⁹. The presence of nonfurocoumarin, umbelliprenin, glycosides of quercetin, luteolin were reported in *Ammi majus* fruits²⁰⁻²¹.

Abdul – Jalil *et al* identified two flavonoids from *Ammi majus* fruit , quercetin and kaempferol . They found that the amount of kaempferol (0.045 %) was higher than quercetin(0.036 %).²² The essential oil extracted from fruits contained high boiling hydrocarbons 1.34%, dipiperitone 10% , unsaturated cyclic terpeniols 15% and a mixture of furocoumarins 60%.²³

Hussain *et al* investigated the fatty acids constituents of *Ammi majus* oil . A total of 18 different components were identified and quantified. Methyl ester of linoleic acid was found in high concentration 9.00%, followed by methyl ester of oleic acid 5.60%, palmitic acid 3.98% and linolenic acids 1.42% . The concentration of the rest identified fatty acids (hexanoic acid, carylic acid capric acid, lauric acid, myristic acid, pentadecanoic acid, palmitic acid, margaric acid, stearic acid, elaidic acid, arachidic acid, behenic acid, tricosnoic acid, tetracosanoic acid) were less than 1%.²⁴

Pharmacological effects

Effects on psoriasis , vitiligo and tinea versicolor

Numerous studies have assessed the efficacy of Fructus *Ammi majus* and xanthotoxin for the treatment of vitiligo, psoriasis, and hypopigmentation tinea versicolor.^{6-8, 25-35}

Experimentation with *Ammi majus* extracts was started in Egypt by El Mofti.^{8, 25} This followed by the work of Sidi and Bourgeois who used *Ammi majus* Linn, in six patients with vitiligo, five men and one woman. Their ages were from 30 to 50 years. *Ammi majus* Linn was used (a) by oral administration, (b) by local topical application at

the affected sites followed by sun or ultraviolet lamp exposure, or, (c) by a combination of (a) and (b). Three of patients were subjected to the combined treatment, two only to topical treatment and one to treatment by mouth for 5 months, and then to the combined treatment. The repigmentation appeared in all patients as pigmented minute macules with hair follicles in their center. These macules were distributed over the leukodermic plaques and increased progressively in size until they joined, forming larger islands. This was particularly distinct in the lesions on the trunk and on the extremities. On the face the repigmentation developed more rapidly and appeared to be progressing more from the periphery towards the center.³⁰

Many clinical trials were carried out to investigate the efficacy of *Ammi majus* in vitiligo. Patient with leukoderma took oral *Ammi majus* powdered fruits with exposing the affected patches to direct sunlight for 1 hour developed symptoms of itching, redness, oedema, vesiculation and oozing in the leukodermic patches. Within few days, the affected skin gradually started to display deep brown pigmentation.⁵

In two small group of patients (eight patients each) with leukoderma treated with oral (0.05 g of *Ammi majus* three time daily) or liniment 1 g/100 ml, applied to the skin, with daily exposure of leukodermic areas to the sun for 0.5 hour or to UV light for 2 minutes, gradually increasing to 10 minutes, the leukodermic skin areas were inflamed and vesiculated, and the leukodermic areas began to show normal pigmentation.⁷

However *Ammi majus* and its furanocoumarins constituents showed good results in many other clinical studies, 70% of the patients treated with an oral dose of 0.6 mg/kg bw of xanthotoxin 2 hours before exposure to sunlight three times per week with calcipotriol ointment in a randomized double-blind study, showed significant improvement.³¹

Xanthotoxin with exposure to either UV-A or UV-B radiation for the treatment of plaque psoriasis in 100 patients appeared effective in reducing the number of plaques.³² Oral administration of 0.6 mg/kg bw of xanthotoxin with two UV-A radiation dosage regimens was used for treatment of patients with moderate-severe chronic plaque psoriasis.

42% of patients were clear 1 year after treatment and the treatment regimens were well tolerated.³³ Many other similar results were obtained in assessment of *Ammi majus* and its furanocoumarins in the treatment of psoriasis, vitiligo and tinea versicolor by many authors.^{7, 28, 34-35}

Other pharmacological effects

Furocoumarins have bactericidal, fungicidal, insecticidal, larvicidal, molluscicidal, nematocidal, ovicidal, viricidal and herbicidal activities.^{3, 36} *Ammi majus* coumarins were evaluated for antiviral effects against two mammalian viruses, HSV-1 and VSV. The antiviral activity was determined by means of the end titration technique that depends on the ability of plant extract dilutions to inhibit the produced cytopathogenic effect. *Ammi majus* coumarins exerted antiviral activity against vesicular stomatitis virus (VSV) in a concentration-dependent manner at complete non-toxic concentration range 10-100 µg/ml. *Ammi majus* coumarins found to have no reliable antiviral activity against herpes simplex virus (HSV).⁹ A dose of 400 mg/kg body weight of a hot aqueous extract and 15.0 mg/kg bw of petroleum ether extract of the *Ammi majus* fruits daily for six days reduced the *Schistosoma mansoni* worm burden in mice by 49.3–72.3%.¹⁸ Mustafa and Al-Khazraji investigated the effects of the extracts *Ammi majus* against larval stage of *Culex pipiens molestus* Forskal. *Ammi majus* L. caused high mortality to the larvae after 7 days of treatment.³⁷ Acetone and 95% ethanol extract of *Ammi majus* inhibited the growth of the *Neurospora crassa* fungi *in vitro*.³⁸ *Ammi majus* coumarins were evaluated for anti-inflammatory activity by the carrageenan induced rat paw edema method. They possessed anti-inflammatory effects at a dose of 0.01 mg/100 g.⁹

Contraindications and adverse effects

A. majus L. is contraindicated in diseases associated with photosensitivity, cataract, invasive squamous-cell cancer, known sensitivity to xanthotoxin (psoralens), and in children under the age of 12 years. The fruits are also contraindicated in pregnancy, nursing, tuberculosis, liver and kidney diseases, human immunodeficiency virus (HIV) infections and other autoimmune diseases.^{9, 39-40}

Patients, after the first exposures, developed bullous reactions of more or less severe but in constant degree similar to burns, nervousness and insomnia,

nausea and gastric burning.³⁰ However, itching, edema, hypotension, vertigo, depression, painful blistering, burning and peeling of the skin, pruritus, freckling, hypopigmentation, rash, cheilitis and erythema were also recorded with xanthotoxin therapy.³⁹ Phototoxic dermatitis and allergic rhinitis and contact urticaria due to exposure to the fruits were recorded.⁴¹⁻⁴² There are also reports of toxicosis by photosensitizing furocoumarins contained in *Ammi majus* seeds in many animal species. In a herd of pigs suffered simultaneous intoxications by ergot alkaloids from *Claviceps purpurea* sclerotia and furocoumarins from *Ammi majus* seeds. Nervous signs were first observed 5-7 days after the initiation of feeding. These signs were followed by cutaneous irritation. Snout ulcers, eyelid edema, and conjunctivitis were recorded in several piglets. Ten days after the start of feeding, 8 abortions were observed. Many of the sows that were nursing piglets developed udder edema and teat cracking. Dermal lesions were observed in most of the animals with unpigmented areas in the skin. Examination of impurities in the suspected wheat indicated the presence of 2.2% of *A. majus* seeds and 0.14% of *C.purpurea* sclerotia. The quantitative analysis indicated the presence of 3.2 g xanthotoxin and 0.65 g bergaptene/100 g *Ammi majus* seeds and 0.73 g ergot alkaloids (expressed as ergonovine) per 100g, of *C. purpurea*.^{18, 42-44}

Dosage

Fructus *Ammi majus* was used as 0.02–0.04 g daily orally in divided doses, xanthotoxin 0.25–0.7 mg/kg bw.^{4,6,8, 28-29}

II-*Ammi visnaga*

Synonyms : *Daucus visnaga* L., *Selinum visnaga* E.H.L. Krause, *Sium visnaga* Stokes, *Visnaga daucoides* Gaertn.⁴⁵⁻⁴⁷

Common names : English: Pick-tooth, Tooth pick, Bishop's weed, Arabic: Khella, Khella baladi

Traditional uses : The fruits of *Ammi visnaga* were used in the treatment of mild anginal symptoms. As supportive treatment of mild obstruction of the respiratory tract in asthma, bronchial asthma or spastic bronchitis, and postoperative treatment of conditions associated with the presence of urinary calculi. Treatment of gastrointestinal cramps and painful menstruation. Internally as an emmenagogue to regulate menstruation, as a diuretic, and for treatment of vertigo, diabetes and kidney stones.¹⁰

Physico-chemical constants^{10, 46}: loss in weight on drying at 105°C: 4.60%, total ash: 9.4%, acid insoluble ash: 0.6%, and water soluble ash: 2.9%. Extractive value in different solvents (%): petroleum ether: 3.40, chloroform (60-80°C): 6.10, absolute ethanol: 11.10 and ethanolic water extract: 19.50.

Chemical constituents

Ammi visnaga contained -pyrones (furanochromone up to 4%), the principal compounds being khellin (0.3–1.2%), visnagin (0.05–0.30%), khellinol, ammiol, khellol and khellinin. *Ammi visnaga* also contained fixed oils (up to 18%) and coumarins (0.2–0.5%), the main one being the pyranocoumarin visnadin (0.3%).⁴⁶⁻⁵⁰ The hydrodistillation of *Ammi visnaga* yielded 1.3% of yellowish oil. Twenty one components were identified representing 97.3% of the essential oil. These compound included 2,2- dimethylbutanoic acid (30.1%), isobutyl isobutyrate (14.0%), croweacin (12.2%), linalool (12.1%), bornyl acetate (7.3%), thymol (6.0%), -thujene (1.5%), 3-methylpentenol (2.5%), -myrcene (0.1%), methylbutyl 2-methylbutanoate (1.2%), -isophorone (3.8%), 2-nonyne (1.2%), hexenyl isobutanoate (1.6%), endo-fenchyl acetate (0.2%), geranyl acetate (1.2%), lavandulyl acetate (1.2%), citronellyl propionate (0.6%), neryl isobutanoate (0.1%), lavandulyl 2- methylbutanoate (0.1%), and -damascone (0.1%), Z,E)-farnesal (trace).⁵¹

Eleven flavonols have been isolated from the aerial parts of *Ammi visnaga* L. from which four aglycones, four monoglycosides, two diglycosides and one triglycoside. The flavonoid aglycones were distributed into one hydroxylated (quercetin) and three methoxylated (rhamnetin, isorhamnetin and rhamnazin). The monoglycosides included three 3-O-glucosides respectively linked to rhamnetin, isorhamnetin and rhamnazin and one 7-O-glucoside of isorhamnetin. The two diglycosides were 3-O-rutin of quercetin and isorhamnetin while the single trioside was quercetin 7,3,3'-O-triglucoside.^{49, 52-54}

Pharmacokinetic studies

The plasma concentration of visnagin after oral dose reached the maximum level of 3270.72 ng/mL at 0.33 h and decreased to below limit of quantitation (1.0 ng/mL) after 12 h. For intravenous administration, the maximum concentration of

visnagin was 1635.76 ng/mL at 0 h. Visnagin at a dose of 10 mg/kg (in 2% ethanol and 10% PEG 200) was completely absorbed (oral bioavailability, F=100.71%). The half-lives of 0.79 and 0.61 was recorded in oral and intravenous administration respectively. The volume of distribution (Vd) of visnagin was 0.86 L, which is suggestive of the distribution into extracellular fluids in the body.⁵⁵

Pharmacological effects

Antimicrobial effects

The antimicrobial effects of the ethanolic and aqueous extract of *Ammi visnaga* were tested against eight pathogenic microorganisms *Staphylococcus aureus*, *Leuconostic mesonstroide*, *Enterococcus faecalis*, *Escherichia coli*, *Pseudomonas aeruginosa*, *Klebsiella pneumoniae*, *Candida tropicans* and *C. albicans*. The most active extract against Gram-positive bacteria was ethanol extract with a minimal inhibitory concentration (MIC) value of (5mg/ml) against *Enterococcus faecalis*. In addition, the same extract exerted antimicrobial activity against the Gram-negative bacteria *Escherichia coli*, *Klebsiella pneumoniae* with an MIC value of 12.5mg/ml. In yeast a high concentration of extract was needed to cause inhibition.⁵⁶

The essential oil of *Ammi visnaga* was tested against *Escherichia coli* ATCC 25922, *Escherichia coli*, *Staphylococcus aureus* ATCC 43300, *Staphylococcus aureus*, *Pseudomonas aeruginosa* ATCC 27853, *Pseudomonas aeruginosa*, *Enterobacter aerogenes*, *Klebsiella pneumoniae*, and *Morganella morganii*. The essential oil exhibited the best antibacterial activity against *Escherichia coli* ATCC 25922, *Escherichia coli*, *Staphylococcus aureus* ATCC 43300 and *Pseudomonas aeruginosa* ATCC 27853, the diameter of the inhibitory zones were 29, 25, 25, 25 mm respectively⁵¹. Ethanol extract of *Ammi visnaga* fruits (at a dilution of 1:40) inhibited the growth of *Mycobacterium tuberculosis* H37RVTMC 102⁵⁷. An aqueous extract of the fruits, 2–10 mg/ml inhibited growth and aflatoxin production of *Aspergillus flavus*, the effects were dose-dependent⁵⁸. The aqueous and hydroalcoholic extract of seed and stem of *Ammi visnaga* showed a good antibacterial activity against *Streptococcus mutans*, *Streptococcus salivarius* and *Streptococcus sanguis* oral pathogens⁵⁹.

Cardiovascular effects

Ammi visnaga induced relaxation of smooth muscle, including that of the ureter and coronary arteries, in a variety of animal species⁶⁰. Durate *et al* found that visnadine caused nonspecific inhibition of vascular smooth muscle. It was selectively inhibited the contractile response in the rat isolated aortic ring and portal vein segment. On the other hand, intravenous administration of visnagin decreased blood pressure with no significant changes on the heart rate⁶¹⁻⁶³. A chloroform, and methanol extract (1mg/ml) of the fruits inhibited the potassium chloride induced contractions of the rabbit guinea-pig aorta in vitro⁶⁴⁻⁶⁶. Visnadin, 60.0 µg/ml or 120.0 µg/ml, increased coronary blood flow in isolated guinea-pig hearts by 46% and 57% respectively⁶⁶. Samidin and khellol glucoside induced positive inotropic effects on heart⁶⁷.

In coronary vasospasm and myocardial ischaemia induced in dogs by daily intramuscular injections of vasopressin, visnadin, dihydrosamidin, khellin and samidin effectively normalized the electrocardiogram when given in a dose of 4.7 mg/kg bw per day intramuscularly for 7 days⁶⁷. Immediately after the rapid intravenous administration of 20-30 mg of khellin to the dogs, the blood pressure drops to about 50 mm Hg., the heart beats considerably slower, and the respiration is momentarily arrested. The entire effect lasts for only a short time, within a minute or two⁶⁸. According to the results obtained by different researchers, khella seems to improve blood supply to smooth muscles and makes myocardial metabolism more efficient. It dilated the coronary vessels, and increased the capacity of the heart without increasing the heart rate or affecting blood pressure⁶⁹.

A clinical trial of khellin in 38 cases of angina pectoris and in 8 cases of coronary thrombosis was performed. Continuous treatment, by the oral or intramuscular routes or by both, gave favourable results in 35 out of 38 cases of angina pectoris. Continuously administration of khellin for several weeks to eight patients after coronary thrombosis appeared favourable⁶⁷.

A clinical study was carried out on 20 non-obese, normolipaeamic male subjects to determine the effects of orally administered 50 mg khellin four times daily for 4 weeks on the plasma lipids. Plasma total cholesterol and triglyceride remained

unchanged, but high-density-lipoprotein cholesterol concentration was significantly elevated during the treatment and till one week after cessation of treatment⁷⁰. In a comparison with glyceryl trinitrate, khellin (3 ml. containing 150 mg. of khellin, alcoholic extract standardized to contain 50 mg/ml) was used in twelve patients for prevention of angina of effort and the electrocardiographic changes that may accompany it. Khellin was less potent but longer acting than glyceryl trinitrate, and it did not cause any unpleasant side effects⁷¹.

Treatment of vitiligo

A double-blind, placebo-controlled study of 60 people indicated that the combination of oral khellin (which is the main constituent of *Ammi visnaga*) and natural sun exposure caused repigmentation in 76.6% of the treatment group, in comparison, no improvement was seen in the control group receiving sunlight plus placebo⁷². A subsequent placebo-controlled study of 36 patients of vitiligo, showed that a topical khellin gel plus UVA caused repigmentation in 86.1% of the treated cases, as opposed to 66.6% in the placebo group⁷³.

Smooth muscle relaxant effects

Durate et al found that visnadine caused nonspecific inhibition of vascular smooth muscle. It was selectively inhibited the contractile response in the rat isolated aortic ring and portal vein segment⁶¹⁻⁶³. Aqueous extract of *Ammi visnaga* seeds induced relaxant effect on contractibility of small intestine of rabbit⁷⁴. *Ammi visnaga* induced relaxation of smooth muscle, including that of the ureter and coronary arteries, in a variety of animal species⁶⁰. Khella's antispasmodic properties are also useful to treat asthma attacks. During the 1950s, research into khella's usefulness as an asthma treatment led to the creation of many asthma medications containing khellin and visnagin⁶⁹.

Prevention of urolithiasis

Ammi visnaga was investigated for the preventive effect of kidney stone formation. In cell culture experiments, it was found that *Ammi visnaga* and its compounds (khellin and visnagin) protected cell damage from calcium oxalate crystals. In addition, *Ammi visnaga* and its compounds prevented calcium oxalate crystals formation in stone forming rats by increasing the urinary pH and citrate concentration along with a decrease of urinary oxalate. The calcium oxalate crystals

deposition in the rat kidneys was significantly decreased in the group of rats receiving *Ammi visnaga* and its compounds⁵⁵.

Antioxidant effects

The antioxidant activity of the butanolic extract of *Ammi visnaga* was determined by 2,2-Diphenyl-1-picryl-hydrazyl (DPPH) method. The butanolic extract of *Ammi visnaga* was markedly quenched the DPPH radical by 78.7% at a concentration of 200 µg/ml⁷⁵.

Contraindications and adverse effects

To minimize photosensitivity, the exposure to sun or other sources of ultraviolet light should be avoided during treatment with *Ammi visnaga* and its components. Long term use or overdose of the drug can lead to queasiness, dizziness, loss of appetite, headache, sleep disorders and with very high dosage (corresponding to over 100 mg khellin), it caused reversible elevation in the levels of liver enzymes⁷⁶⁻⁷⁷. Ethanolic extract of *Ammi visnaga* was free from mutagenic effect, it also inhibits the mutagenic effects of ethyl methanesulfonate, 2-amino-anthracene, and benzopyrene in *S. typhimurium*⁷⁸.

Dosage

Average daily dose: Fructus *Ammi Visnaga* 0.05–0.15 g.^{1,46}

References

1. Ramadan, S. *Ammi majus* plant. *Hamdard* 1982; 25 (1-4):32-35.
2. Chakravarty HL. *Plant wealth of Iraq*, 1st ed. Baghdad Botany Directorate, Ministry of Agriculture and Agrarian, Republic of Iraq 1976: pp 11,27.
3. Joy PP, Thomas J, Mathew S and Skaria M. *Medicinal plants*. Kerala Agricultural University, India 1998.
4. *Egyptian Pharmacopoeia*, General Organization for Government Printing, Cairo 1972, 32.
5. Hakim RE. Rediscovery of a treatment for vitiligo. *Clio Medica* 1969; 4:277–289.
6. El-Mofty AM. A preliminary clinical report on the treatment of leucoderma with *Ammi majus* Linn. *J Egypt Med Assoc* 1948; 31:651–665.
7. Fahmy IR and Abu-Shady H. The isolation and properties of ammoidin, ammidin and

- majudin and their effect in the treatment of leukoderma. Q J Pharm Pharmacol 1984; 21:499-503.
8. El-Mofty AM. Further study on treatment of leukoderma with *Ammi majus* Linn. J R Egypt Med Assoc 1952; 35:1-19.
 9. Selim YA , and Ouf NH. Anti-inflammatory new coumarin from the *Ammi majus* L. Medicinal Chemistry Letters 2012; 2:1-4.
 10. WHO monographs on selected medicinal plants, Vol 3. WHO Library Cataloguing in Publication Data .WHO 2007 pp 23-32.
 11. Ubramanian P, Rajan S and Kumar S. Physico-chemical profile of *Ammi majus*. Ancient Science of Life 1996; XVI : 142 - 147.
 12. Blazek ZA . Pharmacognosy of the plant parts of *Ammi majus* L. Farm Obz, Ch 1966;35(1): 495-509.
 13. Eisenreichoa E, Buckova L and Tomko J. Contents of *Ammi majus* L substances. Farm Obz Ch 1980; 49(11): 503-506.
 14. Ekiert H, and Gomolka E. Coumarin compounds in *Ammi majus* L. callus cultures. Pharmazie 2000; 55:684-687.
 15. Abu-Mustafa EA and Fayez MBE. Natural coumarins. I. Marmesin and marmesinin, further products from the fruits of *Ammi majus* L. Journal of Organic Chemistry 1961; 26:161-166.
 16. Hilal SH and Haggag MY. A thin-layer chromatography (TLC)-colorimetric assay of furocoumarins. Egyptian Journal of Pharmaceutical Sciences 1975; 16:495-499.
 17. Elgamal MHA , Shalaby NMM , Duddeck H and Hiegemann M. Coumarins and Coumarin glycosides from the fruits of *Ammi* species. Phytochemistry 1993 ; 3(34) 819-823.
 18. Abdulla WA.. Preliminary studies on the anti-schistosomal effect of *Ammi majus* L. Egyptian Journal of Bilharziasis 1978; 4:19-26.
 19. Ivie GW. Linear furocoumarins (psoralens) from the seed of Texas *Ammi majus* L. (bishop's weed). Journal of Agricultural and Food Chemistry 1978; 26:1394-1403.
 20. Singab ANB. Acetylated flavonol triglycosides from *Ammi majus* L. Phytochemistry 1998; 49:2177-2180.
 21. Anonymous. The wealth of India – Raw Materials. Vol I , Council of Scientific Industrial Research, New Delhi 1985.
 22. Abdul-Jalil TZ , Saour K and Nasser A. Phytochemical study of some flavonoids present in the fruits of two *Ammi* L. species wildy grown in Iraq. Iraqi J Pharm Sci 2010; 19(1):48-57.
 23. Ashraf M, Ahmad R and Bhatti MK. Studies on the essential oil content of the Pakistani species of the family Umbelliferae. Part XXX *Ammi majus* seed oil. Park J Sci Ind Res 1979;22(5):255-257.
 24. Hussain I, Khan S , Khan MI , Ur Rehma I and Ahmed M . Investigation of fatty Acid composition of *Ammi majus* seed oil by gas chromatography mass spectrometry. J Chin Chem Soc 2012; 59 :1-4 .
 25. El-Mofty AM. A preliminary clinical report on the treatment of leukoderma with *Ammi majus* Linn. J Egypt Med Assoc 1984; 31:651-665.
 26. Parrish JA. Photochemotherapy of psoriasis with oral methoxsalen and long wave ultraviolet light. N Engl J Med 1974; 291:1207-1211.
 27. El-Mofty AM and El-Mofty M. Psoralen photochemotherapy in contrast to chemotherapy of psoriasis. Med J Cairo Univ 1980 ;48:71-83.
 28. El-Mofty AM, El-Sawalhy H and El-Mofty M. Clinical study of a new preparation of 8-methoxypsoralen in photochemotherapy. Int J Dermatol 1994; 33:588-592.
 29. El-Mofty AM, El-Sawalhy H and El-Mofty M. Photochemotherapy in the treatment of post tinea versicolor hypopigmentation. Med J Cairo Univ 1995; 61(4):632-637.
 30. Sidi E and Bourgeois J. The treatment of vitiligo with *Ammi majus* Linn . J Invest Dermatology 1951: 391-395.
 31. Parsad D, Saini R, and Verma N. Combination of PUVAsol and topical calcipotriol in vitiligo. Dermatology 1998; 197:167-170.
 32. De Berker DA. Comparison of psoralen-UVB and psoralen UVA photochemotherapy in the treatment of psoriasis. Journal of the American Academy of Dermatology 1997; 36:577-581.
 33. Collins P. 8-MOP PUVA for psoriasis: a comparison of minimal phototoxic dose-based regimen with a skin-type approach. British Journal of Dermatology 1996; 135:248-254.
 34. Kavli G and Volden G. Phytophotodermatitis. Photodermatology 1984; 1:65- 75.

35. Becker SW. Psoralen phototherapeutic agents. *Journal of the American Medical Association* 1967; 202:422–424.
36. Duke JA. Bishops weed (*Ammi majus* L., Apiaceae). *Econ Bot* 1988; 42 (3):442-445.
37. Mustafa MA and Al-Khazraji A. Effect of some plant extracts on the *Culex pipiens molestus* Forskal larvae. *Iraqi Journal of Veterinary Science* 2008 ; 22(1): 9-12.
38. Kubas J. Investigations on known or potential antitumoral plants by means of microbiological tests. Part III. Biological activity of some cultivated plant species in *Neurospora crassa* test. *Acta Biologica Cracoviensa, Series Botanica* 1972; 15:87–100.
39. Lacy C . *Drug information handbook*. Lexicomp, Hudson, OH 2006; 6.
40. Wagner H and Wisenauer ML. *Phytotherapie. [Phytotherapy.]*. Gustav Fischer, Stuttgart 1995.
41. Ossenkoppele PM, van der Sluis WG, and van Vloten WA. Fototoxische dermatitis door het gebruik van de *Ammi majus*-vrucht bij vitiligo. [Phototoxic dermatitis following the use of *Ammi majus* fruit for vitiligo]. *Nederlands Tijdschrift voor Geneeskunde* 1991; 135:478–480.
42. Kiistala R , Makinen-Kiljunen S , Heikkinen K , Rinne J and Haahtela T. Occupational allergic rhinitis and contact urticaria caused by bishop's weed (*Ammi majus*). *Allergy* 1999; 54:635–639.
43. Odriozola E. Fotosensibilización y queratoconjuntivitis en rumiantes por consumo de semillas de falsa viznaga (*Ammi majus* L.). *Vet Argent* 1984; 1:684-688.
44. Shlosberg A, Egyed M, Eilat A. The comparative photosensitizing properties of *Ammi majus* and *Ammi visnaga* in goslings. *Avian Dis* 1974; 18:544-550.
45. *African pharmacopoeia*. Vol.1. Lagos, Organization of African Unity, Scientific, Technical and Research Commission 1985.
46. *Encyclopedia of medicinal plants in UAE* . Health Authority Abu Dhabi , Zaied Center for Traditional Medicine and Herbs Researches 2005 pp 15-20.
47. Abou-Mustafa EA. A further contribution to the -pyrone constituents of *Ammi visnaga* fruits. *Planta Medica* 1990; 56:134.
48. Martelli P. Rapid separation and quantitative determination of khellin and visnagin in *Ammi visnaga* (L.) Lam. fruits by high-performance liquid chromatography. *Journal of Chromatography* 1984; 301:297–302.
49. Franchi GG. High-performance liquid chromatography analysis of the furanochromones khellin and visnagin in various organs of *Ammi visnaga* (L.) Lam. at different developmental stages. *Journal of Ethnopharmacology* 1985; 14:203–212.
50. El-Domiaty MM. Improved high-performance liquid chromatographic determination of khellin and visnagin in *Ammi visnaga* fruits and pharmaceutical formulations. *Journal of Pharmaceutical Sciences* 1992; 81:475–478.
51. Khalfallah A , Labeled A , Semra Z, Al Kaki B and Kabouche R. Antibacterial activity and chemical composition of the essential oil of *Ammi visnaga* L. (Apiaceae) from Constantine, Algeria. *Int J Med Arom Plants* 2011; 193: 302-305.
52. Kabouche Z and Jay M. Flavonols and antioxidant activity of *Ammi visnaga* L. (Apiaceae). *Rec Nat Prod* 2011; 5(1): 52-55.
53. Cisowski W. Flavonoids of *Ammi visnaga*. Lam. Fruits. *Pol J Chem* 1986; 60: 77-84.
54. Saleh AM. Comparative study of the flavonoids of some local members of the Umbelliferae. *Phytochemistry* 1983; 22: 1417-1420.
55. Vanachayangkul P. *Ammi visnaga* L. for prevention of urolithiasis. PhD thesis , Florida University 2008 .
56. Ghareeb AM, Zedan TH and Gharb LA .Antibacterial and antifungal activities of *Ammi visnaga* extracts against pathogenic microorganisms. *Iraqi Journal of Science* 2011; 52(1) : 30-36.
57. Grange JM, Davey RW. Detection of antituberculous activity in plant extracts. *Journal of Applied Bacteriology* 1990; 68:587–591.
58. Mahmoud A. Inhibition of growth and aflatoxin biosynthesis of *Aspergillus flavus* by extracts of some Egyptian plants. *Letters in Applied Microbiology* 1999; 29:334–336.
59. Semyari1 H, Owlia P, Farhadi S and Saeed Tabrizi M. Evaluation of antimicrobial effect of *Ammi visnaga* against oral streptococci. *Journal of Microbiology and Antimicrobials* 2011; 3(5): 126-129.

60. Quart SK. J Pharm Pharmacol 1930; 4: 25.[Altinterim, B . Hiltan tohumunun (*Umbelliferae, Ammi visnaga* L.) düz kaslar üzerine etkisi. Nev ehir Üniversitesi Fen Bilimleri Enstitü Dergisi 60-64.
61. Durate J, Perez-Vizcaino F, Torres AI, Zarzuelo A, Jimenez J and Tamargo J. Vasodilator effects of visnagin in isolated rat vascular smooth muscle. Eur J Pharmacol 1995; 286 (2) : 115- 122.
62. Durate J, Vallejo I , Perez -Vizcaino F , Jimenez R, Zarzuelo A and Tamargo J. Effects of visnadine on rat isolated vascular smooth muscles. Planta Med 1997; 63 (3):233-236.
63. Durate J, Torres AI and Zarzuelo A. Cardiovascular effects of visnagin on rats. Planta Med 2000; 66 (1) : 35-39.
64. Rauwald HW, Brehm H and Odenthal KP. Screening of nine vasoactive medicinal plants for their possible calcium antagonist activity. Strategy of selection and isolation for the active principles of *Olea europaea* and *Peucedanaum ostruthium*. Phytotherapy Research 1994; 8:135-140.
65. Rauwald HW, Brehm H and Odenthal KP. The involvement of Ca²⁺ channel blocking mode of action in the pharmacology of *Ammi visnaga* fruits. Planta Medica 1994; 60:101–105.
66. Erbring H, Uebel H, Vogel G and Chemie Z. Pharmakologie und Toxicologie von Visnadin. [Chemistry, pharmacology, and toxicology of visnadine.] Arzneimittelforschung 1967;17:283-287.
67. Galal EE, Kandil A, Latif MA. Evaluation of cardiac inotropism of *Ammi visnaga* principles by the intra-ventricular technique. Journal of Drug Research of Egypt 1975; 7:45–57.
68. Anrep GV , Barsoum GS , Kenawy MR , and Misrahy G. *Ammi visnaga* in the treatment of angina syndrome. Gazette of the Faculty of Medicine, Cairo 1945; 13, 39.
69. Altinterim B. Hiltan tohumunun (*Umbelliferae, Ammi visnaga* L.) düz kaslar üzerine etkisi. Nev ehir Üniversitesi Fen Bilimleri Enstitü Dergisi 60-64.
70. Harvengt C, and Desager JP. HDL-cholesterol increase in normolipaemic subjects on khellin: a pilot study. International Journal of Clinical Pharmacology Research 1983; 3:363–366.
71. Dewar HA and Grimson TA . Khellin in the treatment of angina of effort. Br Heart J 1950; 12: 54-60.
72. Abdel-Fattah A, Aboul-Enein MN and Wassel GM. An approach to the treatment of vitiligo by khellin. Dermatologica 1982;165:136–140.
73. Orecchia G, Sangalli ME, Gazzaniga A. Topical photochemotherapy of vitiligo with a new khellin formulation: preliminary clinical results. J Dermatol Treat 1998;9:65–69.
74. Jawad AAD, Khuon OS and Ali NA. Spasmolytic activity of *Ammi visnaga* seeds on isolated rabbit jejunum. Basrah Journal of Scienc 2006; 24(1): 47-58.
75. Kabouche Z and Jay M. Flavonols and antioxidant activity of *Ammi visnaga* L. (Apiaceae). Rec Nat Prod 2011; 5(1): 52-55.
76. Blumenthal M (ed). The complete German Commission E monographs. Austin, TX, American Botanical Council, 1998.
77. PDR for herbal medicines , Medical Economic Co. Montvale , New Jersey, 1998, p639.
78. Schimmer O and Rauch P. Inhibition of metabolic activation of the promutagens, Benzo [a]pyrene, 2-aminofluorene and 2-aminoanthracene by furanochromones in *Salmonella typhimurium*. Mutagenesis 1998; 13:385–389.