
Research Article



ISSN Print 2231 – 3648
 Online 2231 – 3656

Available Online at: www.ijpir.com

**International Journal of
Pharmacy and Industrial
Research**

**EFFECT OF AQUEOUS AND ETHANOL EXTRACTS OF
OCIMUM LAMIIFOLIUM AND *AMARANTHUS DUBIUS* AGAINST
BACTERIA ISOLATED FROM CLINICAL SPECIMEN**

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Abstract

The aim of the study was to screen the antibacterial activities of aqueous and ethanol extracts of the two plants *Ocimum lamiifolium* and *Amaranthus dubius* available in Ethiopia. Crude extract of the leaves with aqueous and ethanol were screened for antibacterial activities against *S. aureus*, *Pseudomonas* spp. and *Escherichia coli*. The in vitro antibacterial activity was performed by agar disc diffusion method. Ethanol and aqueous extract of *Ocimum lamiifolium* and *Amaranthus dubius* revealed an elevated antimicrobial activity against *S. aureus*, *E. coli* and *Pseudomonas* spp. The present study shows that the maximum effect was exhibited by *Ocimum lamiifolium*. The results obtained in the present study suggest that the ethanol and aqueous extracts of *Ocimum lamiifolium* and *Amaranthus dubius* revealed a significant scope to develop a novel antibacterial herbal formulation.

Keywords: *Ocimum lamiifolium*, *Amaranthus dubius*, Crude extracts, Antibacterial effects.

Introduction

Plants are a source of many potent and powerful drugs used medicinally in different countries. Medicinal plants are used by 80% of the world population as the only available medicines especially in developing countries¹. Current research on natural molecules and products primarily focuses on plants since they can be sourced more easily and be selected based on their ethno-medicinal uses². A wide range of medicinal plants parts is used to extract as raw drugs and they possess varied medicinal properties. While some of these raw drugs are collected in smaller quantities

by the local communities and folk healers for local used, many other raw drugs are collected in larger

quantities and traded in the market as the raw materials for many herbal industries³. Plants used for traditional medicine contain a wide range of substances that can be used to treat chronic as well as infectious diseases. Clinical microbiologists have great interest in screening of medicinal plants for new therapeutics⁴. The active principles of many drugs found in plants are secondary metabolites. The antimicrobial activities of plant extracts may reside in a variety of different components, including aldehyde and phenolic compounds⁵. The development of drug resistance

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in human pathogens against commonly used antibiotics has necessitated a search for new antimicrobial substances from other sources including plants⁶. Screening of medicinal plants for antimicrobial activities is important for finding potential new compounds for therapeutic use.

In recent years interest to evaluate plants possessing antibacterial activity for various diseases is growing⁷. Research has been done worldwide on the use of medicinal plants to cause human diseases and extracts of many plant species have been found to be active against many pathogenic bacteria. Aqueous extract of some plants have been reported from time to time to demonstrate antimicrobial activity. The efficacy of various species of medicinal plants against a variety of pathogens has been reported by a number of workers⁸⁻¹⁷.

A wide variety of indigenous and minor crops have been utilized for daily consumption since ancient times. The family Labiateae is one of the largest families, which comprises the larger proportion of medicinal plant species. *Ocimum* is one of the important genera of family Labiateae. *Ocimum* species often referred to as the "king of the herb". *Ocimum lamifolium* is an important medicinal herb belonging to family Lamiaceae. The plant extracts and isolated compounds of *Amaranthus* species have been tested for their bioactivity by various in vitro model systems¹⁸.

Materials and Methods

Plant material

The *Ocimum lamifolium* and *Amaranthus dubius* were collected from different area of Gondar region of Ethiopia in September, 2011. The species were botanically identified.

Preparation of the extracts

The leaves were cleaned with running water and dried first in sun light for seven days and then in an oven at 40°C for about 24 hours. Finally, the dried materials were pulverized into fine powdered substance by a grinding machine. Twenty gram of powder of *Ocimum lamifolium* and *Amaranthus dubius* were weighted with the electric balance and transferred into two separate 100 ml conical flasks. Then 40 ml of ethanol in one flask and 40 ml of distilled water in another was added. The conical flasks were closed by foil paper and put on dark

place for maximum 7 days. The crude ethanol and aqueous extracts were then filtered by passing the extracts through Whatmann No. 1 filter paper and then concentrated under vacuum at 40°C by using a rotary evaporator. The residual extracts were stored in refrigerator at 4°C in small and sterile plastic bottles.

Tested bacteria

Antibacterial activity of spices powder extracts was investigated against two gram-negative and two gram positive bacterial isolates, which were obtained from the Hospital Laboratory of Gondar University, Ethiopia. These include *Pseudomonas spp.*, *Escherichia coli* and *S.aureus*. The tested bacteria were cultured on Nutrient agar at 37°C for 24 h.

Inoculum preparation

Ten ml of distilled water was taken into the screw cap tube and pure colony of freshly cultured bacteria was added into the tube and vortex was done. The OD (optical density) was measured with the colorimeter and microbial population was confirmed to be within in 10^7 ml⁻¹ to 10^8 ml⁻¹. This suspension was used as inoculums.

Antimicrobial assay

The in vitro antibacterial activities of the test samples were carried out by disc diffusion method^{19, 20}. In the disc diffusion method, Muller-Hinton agar was used as culture media and the discs were placed aseptically over the bacterial culture on nutrient agar plates and incubated at 37°C for 24 hours. After incubation for 24 hours, the zones of inhibition around the discs were measured by millimeter scale. Discs were impregnated with each treatment and control was assayed on agar medium plate for *Pseudomonas spp.*, *Escherichia coli* and *S. aureus*. The diameter of zone of inhibition as indicated by clear area which was devoid of growth of microbes was measured to determine the antibacterial activity. The experiment was repeated two times to confirm the reproducible results. Sterile, blank paper discs impregnated with only sterile ethanol and distilled water served as negative control each time. Standard Gentamycin (10µg/disc) was used as positive control for comparison of the antibacterial activity. Minimum Inhibitory Concentration (MIC) value of the extracts of the *Ocimum lamifolium* and

Amaranthus dubius were determined in present study following the serial dilution technique.

Results

The quantity of ethanol and aqueous extracts of *Ocimum lamifolium* and *Amaranthus dubius*

showed in table. 1. The maximum 10.7% and 12.9% was extracted from both the plant using aqueous extraction method. Whereas, least 9.4% and 11% yield weighed from both the plants using ethanol as a solvent.

Table No. 01: Yield of plant extract using ethanol and aqueous extraction methods

Plant Species	Parts used	Extraction Type	% Yield (w/w)
<i>Ocimum lamifolium</i>	Leaf	Ethanol	9.4
		Aqueous	10.7
<i>Amaranthus dubius</i>	Leaf	Ethanol	11
		Aqueous	12.9

Antimicrobial activities

Ethanol and aqueous extracts of *Ocimum lamifolium* were found sensitive to *S. aureus*, *E. coli* and *P. aeruginosa*. Crude ethanol extract produced zone of inhibition 12 mm and 13.5 mm against *S. aureus* and *E. coli* respectively and no zone of inhibition was observed for *P. aeruginosa*. Crude aqueous extract produced zone of inhibition of 13.5 mm, 15.5mm and 13 mm against *S. aureus*, *E. coli* and *P. aeruginosa* respectively (Table 2). However, the aqueous extract exhibited highest zone of inhibition (15.5 mm) against *E. coli*.

Ethanol and aqueous extracts of *Amaranthus dubius* were also found sensitive to *S. aureus*, *E. coli* and *P. aeruginosa*. Crude ethanol extract produced zone of inhibition 8.5 mm, 9 mm and 12 mm against *S. aureus*, *E. coli* and *P. aeruginosa* respectively.. Crude aqueous extract produced zone of inhibition of 10.5 mm against *S. aureus*. No zone of inhibition was observed for *E. coli* and *P. aeruginosa* (Table 2). However, the ethanol extract exhibited highest zone of inhibition (12 mm) against *P. aeruginosa*.

Table No. 02: Antibacterial activity of ethanol and aqueous extracts using agar well diffusion method

Micro organisms	Mean zone of incubation (mm)				Gentamycin	Dist. Water
	<i>Ocimum lamifolium</i>		<i>Amaranthu sdubius</i>			
	E	Aq.	E	Aq.		
<i>S. aureus</i>	12	13.5	8.5	10.5	26	-
<i>E. coli</i>	13.5	15.5	9	-	17	-
<i>P. aeruginosa</i>	-	13	12	-	31	-

E- Ethanol

Aq. - Aqueous

The MIC value was also determined against the all tested bacteria. The MIC value of ethanol extract of *Ocimum lamifolium* was found to be 200mg and 100 mg against both *S. aureus* and *E. coli* respectively. Whereas, the MIC value of aqueous extract showed 200 mg against *S. aureus* and 100

mg against *E. coli* and *P. aeruginosa* each. The MIC value of ethanol extract of *Amaranthus dubius* was found to be 500mg against all tested bacteria and aqueous extract showed 1000mg against *S. aureus* (Table 3).

Table No. 03: MIC of plant extracts against bacterial isolates (mg/ml)

MO	<i>Ocimum lamifolium</i>		<i>Amaranthus dubius</i>	
	E	Aq.	E	Aq.
<i>S. aureus</i>	200	200	500	1000
<i>E. coli</i>	100	100	500	-
<i>P. aeruginosa</i>	-	100	500	-

E- Ethanol

Aq. - Aqueous

Discussion

The present study showed that most of the plant extract have antibacterial activity against some of the common microorganisms of medical importance. This work could justify their traditional use in treatment of different diseases in human and animals. The fact that both ethanol and aqueous extracts of some plants are showing similar efficacy against some species of bacteria could be due to extraction ability of active ingredients responsible for antibacterial activity by the two extraction systems. On the other hand, some plants have shown variable activity by the two extraction methods presumably because of difference in extracting ability of specific active ingredient by the two extraction methods. Lino and Deogracious^{21, 22} also reported difference in antibacterial activity when two extraction methods were used. The present study shows that the maximum effect was exhibited by *Ocimum lamifolium*. These results are in parallel with the previous report on *Ocimum spp.* by²³. However, in the present study the maximum effect of tested plant extracts were showed on *E. coli* followed by *S. aureus* and *P. aeruginosa*. This result is in contradiction with the report by Pankaj and Puroshottam (2011), revealed that *S. aureus* (a Gram-positive bacterium) was observed as most susceptible bacterium as it was inhibited by extracts of *Ocimum spp.*. *Streptococcus pyogenes* and *Salmonella typhi* was found to be resistant to all the extracts tested²⁴. The results of present study support the traditional usage of plant and *Ocimum lamifolium* and *Amaranthus dubius* plant extracts which possess compounds with antibacterial properties that can be used as antibacterial agents in new drugs for the therapy of infectious diseases caused by pathogens and further work may be carried out for pharmacological evaluation.

Conclusion

The present investigation revealed that the extract of *Ocimum lamifolium* and *Amaranthus dubius* leaves have potent antimicrobial activity which explains its use in traditional system of medicines. Hence, *Ocimum lamifolium* and *Amaranthus dubius* can be employed as a source of natural antimicrobials that can serve as an alternative to conventional medicines.

Acknowledgements

Authors are grateful to department of Medical Microbiology, School of Biomedical and Laboratory Sciences, University of Gondar, Ethiopia for provision of facilities required for experiments and support for this work.

References

1. Hashim H, Kamali EL, Mohammed Y. Antibacterial activity and phytochemical screening of ethanolic extracts obtained from selected Sudanese medicinal plants. *Curr. Res. J. Biol. Sci.*, 2(2), 2010, 143-146.
2. Arora DS, Kaur GJ. Antibacterial activity of some Indian medicinal plants. *J. Nat. Med.*, 61, 2007, 313-317.
3. Uniyal SK, Singh KN, Jamwal P, Lal B, 2006. Traditional use of medicinal plants among the tribal communities of Chhota Bhangal. Western Himalaya. *J. Ethnobiol. Ethnomedi.*, 2, 2006, 1-14.
4. Periyasamy A, Rajkumar, Mahalingam K. Phytochemical screening and antimicrobial activity from five Indian medicinal plants against human pathogens. *Middle-East J. Sci. Res.*, 5(6), 2010, 477-482.
5. Lai PK, Roy J. Antimicrobial and chemo preventive properties of herbs and spices. *Current Medi. Chem.*, 11, 2004, 1451-1460.
6. Erdogru OT, 2002. Antibacterial activities of some plant extracts used in folk medicine. *Pharm. Biol.*, 40, 2002, 269-273.
7. Clark, A. M. and C. D. Hufford. Development of novel prototype antibiotics for opportunistic immunodeficiency syndrome: Human medicinal agents from plants. American chemical Society (ASC symposium series 534), Washington, D.C., 1993, 228-241
8. Bachir, R. G. and M. Beenali. Antibacterial activity of leaf essential oil of Eucalyptus globules and Eucalyptus camaldulensis. *A. J. Pharma. and Pharmacol.*, 2(10), 2008, 211-215.
9. Chao, Sc and D. G. Young. Screening of inhibitory action of essential oils on selected bacteria, fungi and viruses. *J. Essential Oil*, 12, 2000, 102-109.

10. Delpuis, P. J., K. Stanich and M. G. GrardBand .Antimicrobial activity of individual and mixed fraction of dill, Coriander and Eucalyptus oils.*I. J. Food Microbial.*, 74, 2002, 101-109.
11. Singh, G., P. H. Marimutha and C. Catalan. Antimicrobial and antioxidant potential of essential oil and acetone extract of *Myristicafragrans*Houtt (aril part).*J. Food Science*, 70, 2005, 141-148.
12. Somchit, M. N. I., I. Relzal,Flyshanurand and A. R. Mutalin. In vitro antimicrobial activity of ethanol and water extract of *Cassia alata*. *Journal of Ethnopharmacology*, 84, 2002, 1-4.
13. Gullnce, M. M., D. Sokmeu, G. Daferera, H. Agar, N. Orkan, M. P. Kartal, A. Sokemen and E. Sahin. In vitro antibacterial antifungal and antioxidant activity of the essential oil and methanol extract of herbal plant and callus culture of *Saturejahortensis* L. *J. Agricul. Food Chemical*, 51, 2003, 3958-3965.
14. Hammer, K. A., C. F. Carson and T. V. Riley. Antimicrobial activity of essential oil and other plant extracts. *J. Applied Microbiol.*, 86, 1999, 985-990.
15. Mahesh, B. and S. Satish. Antimicrobial activity of some important medicinal plant against plant and human pathogen.*World J. Agriculture Sciences*, 4(5), 2008, 839-843.
16. Rajesh, S. and M. N. Khan. Antimicrobial activity of *Lantana camara* root and stem extract against *Staphylococcus aureus*. *I. Res. J. Bioscience and Biotechnology*, 2(2), 2004, 123-126.
17. Rora, C., J. J. Carramifiana, J. Burillo and A. Herrera. In vitro antibacterial activity of essential oil from aromatic plant against selected food borne pathogen.*J. Food Production*, 67(6), 2004, 1252-1256.
18. Z. C. Maiyo, R. M. Ngunel, J. C. Matasyoh and R. Chepkorir. Phytochemical constituents and antimicrobial activity of leaf extracts of three Amaranthus plant species. *African J. Biotechnology*, 9(21), 2010, 3178-3182.
19. Bauer AW, Kirby WM, Sherris JC, Turck M. Antibiotic susceptibility testing by a standardized single disc method. *American J. Clinical Pathology*, 45, 1996, 493-496.
20. NCCLS, 2000. Performance standards for antimicrobial disk susceptibility tests: Approval standard M2-A7, 7th edition. Pennsylvania: Clinical and Laboratory Standards Institute.
21. Lino A, Deogracious O. The in vitro antibacterial activity of *Annonasenegalensis*, *Securidacca longipendiculata* and *Steanotaeniaaraliacea*-Ugandan medicinal plants.*Afr. Health Sci.*, 6, 2006, 31-35.
22. Parekh J, Jadeja D, Chanda S. Efficacy of aqueous and methanol extracts of medicinal plants for potential antibacterial activity, *Turk. J. Biol.*, 29, 2005, 203-210.
23. AnandSagar and Indira Thakur. Antibacterial activity of *Ocimum sanctum* (linn.), *Murrayakoenigii* (linn.) Spreng and *Artemisia vulgaris* (linn.),*Plant Archives*, 12(1), 2012, 377-381.
24. Pankaj Goyal and Purshotam Kaushik. In vitro Evaluation of Antibacterial Activity of Various Crude Leaf Extracts of Indian Sacred Plant, *Ocimum sanctum* L. *B. Microbiol. Res. J.*, 1(3), 2011, 70-78.